

# **Management Plan**

# For Antarctic Specially Protected Area No. 161 TERRA NOVA BAY, ROSS SEA

## Introduction

The ASPA of Terra Nova Bay is a coastal marine area encompassing 29.4 km<sup>2</sup> between Adélie Cove and Tethys Bay, Terra Nova Bay, immediately to the south of the Italian Mario Zucchelli Station (MZS). Terra Nova Bay was originally designated as Antarctic Specially Protected Area through Measure 2 (2003) after a proposal of Italy. CCAMLR considered and approved its designation during CCAMLR XXVI, Hobart 2007. The Management Plan has been revised in 2008, through measure 14 (2008), in 2013 through measure 15 (2013) and in 2019 through measure 7 (2019).

The primary reason for the designation of Terra Nova Bay as an Antarctic Specially Protected Area (ASPA) is its particular interest for ongoing and future research. Long term studies conducted in the last 30 years by Italian and international scientists have revealed a complex array of species assemblages, characterized by unique symbiotic interactions. In this Area, several VME species are also present, above all the Antarctic scallop *Adamussium colbecki* and pterobranchs, and new species continue to be described.

The high ecological and scientific values derived from the diverse range of species and assemblages, together with the vulnerability of the Area to disturbance by scientific oversampling, alien introductions, and direct human impacts arising from increasing activities at the nearby permanent scientific stations are such that the Area requires long-term special protection.

ASPA 161 is listed within the Environmental Domain S and T (Morgan F. *et al.* 2014. Environmental Domains of Antarctica Version 2.0, Final Report) and protects adjacent Important Bird Area 177 Adélie Cove. As a marine area, it is not part of Antarctic Conservation Biogeographic Regions (Resolution 6, 2012).

# 1. Description of Values to be Protected

This coastal marine area is an important area for well-established and long-term scientific investigations that allowed, up to now, collection of an extensive amount of scientific data. During the last 5 years, substantial scientific research carried out in the ASPA led to significant advances in the knowledge of this area, as demonstrated by the great number of publications produced (see References and relevant supporting bibliography).), many of which contributed to refine the lists of species present in the area and their nomenclature, especially thanks to the use of molecular techniques (barcoding and metabarcoding). The site typically remains ice-free in summer, which is rare for coastal areas in the Ross Sea region, making it an ideal and accessible site for research into the near-shore benthic communities of the region. Extensive marine ecological research has been carried out at Terra Nova Bay since 1986/87, contributing substantially to our understanding of the marine communities in this area, and of the effect of katabatic winds on the physical, chemical and biological processes occurring in the water column (Povero *et al.*, 2001).

High diversity at both species and community levels makes this Area of high ecological and scientific value. Studies have revealed a complex array of species assemblages, often co-existing in mosaics (Sarà *et al.*, 1992; 2002; Gambi *et al.*, 1997; Cantone *et al.*, 2000; Ghiglione *et al.*, 2013) and characterized by unique symbiotic interactions (Schiaparelli *et al.*, 2011; 2015; Regoli *et al.*, 2004). There exist assemblages with high species richness and complex functioning, such as the sponge and anthozoan communities, alongside loosely structured, low diversity assemblages. In this area several VME species also occur, above all the Antarctic scallop Adamussium colbecki (Schiaparelli and Linse, 2006) and pterobranchs (Schiaparelli *et al.*, 2004), and new species continue to be described (see section 6). A population of Adélie penguins (Pygoscelis adeliae) is present nearby the Area.

The collected scientific data over the years, allowed the site to serve as reference for the determination of impacts arising from human activities (Berkman and Nigro, 1992; Focardi *et al.*, 1993; Minganti *et al.*, 1995; Bruni *et al.*, 1997; Nonnis Marzano *et al.*, 2000, Lo Giudice *et al.*, 2013), the understanding of inter-annual variability in species dynamics (Cecchetto *et al.*, 2021) and the assessment of predictive models of species occurrences (Peel *et al.*, 2019; Grillo *et al.*, 2022)



# 2. Aims and Objectives

- avoid degradation of, or substantial risk to, the values of the Area by preventing unnecessary human disturbance to the Area;
- allow scientific research on the ecosystem, in particular on the marine species assemblages and long-term monitoring, while ensuring protection from oversampling or other possible human impacts;
- allow other scientific research and support activities provided they are for compelling reasons which cannot be served elsewhere and that will not compromise the values for which the Area is protected;
- prevent or minimise the possibility of introduction of non-native species (e.g. alien plants, animals and microbes) into the Area;
- allow visits for management purposes in support of the aims of the Management Plan.

# **3. Management Activities**

The following management activities are to be undertaken to protect the values of the Area:

- A map showing the location of the Area (stating the special restrictions that apply) shall be displayed, and a copy of this Management Plan shall be kept available, at all the scientific stations located within 50 km of the Area. Information illustrating the location and boundaries with clear statements of entry restrictions is displayed on posters at MZS;
- Buoys, or other markers or structures established for scientific or management purposes shall be secured and maintained in good condition, and removed when no longer necessary;
- Any abandoned equipment or material shall be removed to the maximum extent possible, provided that doing so does not adversely impact on the environment and the values of the Area;
- Visits shall be made as necessary to assess whether the Area continues to serve the purposes for which it was designated and whether management and maintenance measures are adequate;
- National Antarctic Programs are encouraged to consult together to prevent oversampling within the Area.

# 4. Period of Designation

Designated for an indefinite period.

## 5. Maps and Photographs

• Map 1: Terra Nova Bay, Antarctic Specially Protected Area No. 161, bathymetric map.

**Map specifications:** Projection: UTM Zone 58S; Spheroid: WGS84. Bathymetric contour interval 50 m. Land contours and coast derived from 1:50,000 Northern Foothills Satellite Image Map (Frezzotti *et. al.* 2001). Bathymetry within ASPA derived from high resolution sidescan sonar data surveyed by Kvitek, 2002. Bathymetry outside of ASPA supplied by Italian Hydrographic Office 2000. Marine data collected under Terra Nova Bay marine protected area Project (PNRA 1999-2001). **Inset 1:** The location of Terra Nova Bay in Antarctica. **Inset 2:** Terra Nova Bay location map, showing the region covered by Map 1, stations, and sites of nearby protected areas.

## 6. Description of the Area

#### 6 (i) Geographical co-ordinates, boundary markers and natural features

#### General description, borders and coordinates

The Area is situated in Terra Nova Bay, between the Campbell Glacier Tongue and Drygalski Ice Tongue, Victoria Land (Map 1). It is confined to a narrow strip of coastal waters to the south of MZS (Italy), extending approximately 9.4 km in length and generally within 1.5 – 7 km of the shore, comprising an area of 29.4 km2. No marine resource harvesting has been, is currently, or is planned to be conducted within the Area, nor in the immediate surrounding vicinity.

The western boundary of the Area is defined as the mean high water mark along the coastline extending between 74°42′57″S in the north (2.3 km south of MZS) and 74°48′00″S in the south (the southern shore of Adélie Cove), and includes the intertidal zone (Map 1). The northern boundary of the Area is defined as the 74°42′57″S line of latitude, extending from the coast 1.55 km eastward to the 164°10′00″E line of longitude. The boundary position may be recognised near the shore by the presence of a large and distinctive offshore rock in the northernmost cove on the coast south of MZS, which is an unique feature on this stretch of coast. The southern boundary is defined as the 74°48′00″S line of latitude, extending from the coast 3.63 km eastward to the 164°10′00″E line of longitude. The boundary position may be recognized visually as being at the southern shore of the mouth of Adélie Cove, immediately south of a distinctive rocky outcrop at the base of the coastal cliffs. The eastern boundary of the Area is defined as the 164°10′00″E line of longitude extending between 74°42′57″S in the north and 74°48′00″S in the south.



#### Geology

The coastline of Terra Nova Bay is characterised predominantly by rocky cliffs, with large boulders forming occasional 'beaches' (Simeoni *et al.*, 1989). In the sheltered areas, the soft bottom begins at a depth of 20–30 m. The tidal range is 1.5–2 m and pack ice approximately 2–2.5 m thick covers the sea surface for 9–10 months of the year (Stocchino and Lusetti, 1988; 1990). Data available for the summer period suggest that ocean currents in the Area are likely to be slow and to flow generally in a north-south direction. Along the coastline of the Area there are two main coves; the larger Adélie Cove in the south and a smaller cove around 3 km to its north. The sea floor substrate of the smaller consists of pebbles of various sizes, while Adélie Cove is characterised by fine-grained, muddy sediments. The seafloor within the Area is primarily granitic rock, with softer substrates composed of coarse sands or gravels.

#### Marine benthic communities (0-500 m)

Several new taxa have been described from Terra Nova Bay in the framework of PNRA expeditions. A list of species is reported here below, where type species of new genera are marked with an asterisk (\*) and type localities falling within the ASPA boundaries are reported with two asterisks (\*\*). Most taxa having type localities in Terra Nova Bay or Tethys Bay have also been confirmed to occur within the ASPA boundaries.

Species	Phylum	Type locality	Reference
<i>Craspedostauros ineffabilis</i> Trentin, Moschin, Lopes, Custódio & Moro 2022	Heterokontophyta	Terra Nova Bay	Trentin <i>et al.,</i> 2022
<i>Craspedostauros zucchellii</i> Trentin, Moschin, Lopes, Custódio & Moro 2022	Heterokontophyta	Terra Nova Bay	Trentin <i>et al.,</i> 2022
Tethysphytum antarcticum Sciuto, Moschin & Moro, 2021 (*)	Rhodophyta	Tethys Bay	Sciuto <i>et al.,</i> 2021
Thalassolithon adeliense Trentin, Moschin & Moro 2023 (*)	Rhodophyta	Adelie Cove (**)	Trentin <i>et al.,</i> 2023
Vellaria zucchellii Sabbatini, Pawlowski, Gooday & Bowser, 2004	Foraminifera	Tethys Bay	Sabbatini <i>et al.,</i> (1999)
Megapogon schiaparellii Alvizu, Xavier & Rapp, 2019	Porifera	Tethys Bay	Alvizu <i>et al.,</i> 2019
Microxina sarai Calcinai & Pansini, 2000	Porifera	Adelie Cove (**)	Calcinai & Pansini, 2000
<i>lophon terranovae</i> Calcinai & Pansini, 2000	Porifera	Faraglione (**)	Calcinai & Pansini, 2000
Ectyodoryx minuta Calcinai & Pansini, 2000	Porifera	Terra Nova Bay	Calcinai & Pansini, 2000
Microxina lanceolata Calcinai & Pansini, 2000	Porifera	Terra Nova Bay	Calcinai & Pansini, 2000
Crella (Crella) aurantiaca Bertolino, Calcinai & Pansini, 2009	Porifera	Adelie Cove (**)	Bertolino <i>et al.,</i> 2009
Mycale (Aegogropila) denticulata Bertolino, Calcinai & Pansini, 2009	Porifera	Terra Nova Bay	Bertolino <i>et al.,</i> 2009
Amphicteis teresae Schiaparelli & Jirkov, 2016	Annelida	Terra Nova Bay	Schiaparelli & Jirkov, 2016
Amage giacomobovei Schiaparelli & Jirkov, 2021	Annelida	Terra Nova Bay	Schiaparelli & Jirkov, 2021
<i>Lepidepecreella debroyeri</i> Schiaparelli, Chiara, Kilgallen, Scinto & Lörz, 2015	Arthropoda	Terra Nova Bay	Schiaparelli <i>et al.,</i> 2015
<i>Alcyonidium kuklinskii</i> Schwaha, Cometti, Saadi, Cecchetto & Schiaparelli, 2023	Bryozoa	Tethys Bay	Schwaha <i>et al.,</i> 2023

In the supralittoral zone, only cyanobacteria and diatoms colonise the hard substrates, while the intertidal zone (1.5–2.0 m wide) has, in the most sheltered areas, a high coverage of the green alga Urospora penicilliformis and Prasiola crispa (Cormaci et al., 1992b). Below the tidal zone, down to 2–3 m depth, the community is very poor, due to the persistent presence and scouring action of pack ice, and is mainly composed of epilithic diatoms and the amphipod Paramoera walkeri. Immediately deeper, rocks can be fully colonised by the red alga Iridaea cordata (Cormaci et al., 1996), frequently found with Plocamium cartilagineum, to a depth of 12 m (Gambi et al., 1994; 2000a). At this level, large sessile animals such as Alcyonium antarcticum and Urticinopsis antarctica can be occasionally observed, while frequent are the asteroid Odontaster validus and the echinoid Sterechinus neumayeri. Phyllophora antarctica is another red alga forming expanded mats from 12 to 25 m depth, often heavily colonised by sessile organisms, mainly hydroids (Cerrano et al., 2000c, Puce et al., 2002), serpulids and bryozoans (Celleporella antarctica and Harpecia spinosissima). The upper algal belts represent shelter and a food source for diversified and abundant communities of mobile fauna. Numerous invertebrates, such as the polychaete Harmothoe brevipalpa, the gastropod mollusc Laevilittorina antarctica, the crustacean amphipod Paramoera walkeri and the tanaid Nototanais dimorphus feed on these algal species and can be very abundant. On rocky bottoms in deeper layers, the thalloid macroalgal community is absent and only a calcareous crustose coralline alga previously determined as Clathromorphum lemoineanumund and later erroneously assigned to Phymatolithon foecundum by Alongi et al. (2002). Recent molecular studies have questioned the correctness of previous determination and issued new taxa, such as the new genus Tethysphytum and the species Tethysphytum antarcticum (Hapalidiales, Rhodophyta) (Sciuto et al., 2021). From the same group of algae, also the new genus Thalassolithon and the new species Thalassolithon adeliense (Trentin et al., 2023) have been described from Adelie Cove.



The soft bottoms from 20–40 m depth are coarse sands and gravels, where the community is characterised by the bivalve mollusc Laternula elliptica and the polychaete Aglaophamus ornatus (Nephtiidae). The bivalve Yoldia eightsi is abundant in fine-sand sediments especially in Adelie Cove. From this depth range several new other taxa have been recently described, such as the new genus Tethysphytum and the species Tethysphytum antarcticum of non-geniculate coralline algae (Hapalidiales, Rhodophyta) (Sciuto *et al.*, 2021) and, the same group, the new genus Thalassolithon and the new species Thalassolithon adeliense (Trentin *et al.*, 2023), the new, such as the Antarctic diatoms Craspedostauros ineffabilis and Craspedostauros zucchellii (Trentin *et al.*, 2022), the new bryozoan Alcyonidium kuklinskii (Schwaha *et al.*, 2023) and the calcareous sponge Megapogon schiaparellii (Alvizu *et al.*, 2019).

Between 30–70 m, the substrate becomes finer and is completely colonised by the bivalve Adamussium colbecki, the shells of which are colonised by a micro-community comprising mainly forams, bryozoans (Aimulosia antarctica, Arachnopusia decipiens, Ellisina antarctica, Micropora brevissima) and the spirorbid Paralaeospira levinsenii (Albertelli *et al.*, 1998; Ansell *et al.*, 1998; Chiantore *et al.*, 1998; 2000; 2001; 2002; Vacchi *et al.*, 2000a; Cerrano *et al.*, 2001a; 2001b). In this region, large predators such as the gastropod Neobuccinum eatoni and the nemertean Parborlasia corrugatus are frequent. The echinoid Sterechinus neumayeri and the starfish Odontaster validus are still very frequent at all depths on both hard and mobile substrates (Chiantore *et al.*, 2002; Cerrano *et al.*, 2000b). Several unique biotic associations have been described at these depths, e.g. between sponges and other invertebrates (Schiaparelli *et al.*, 2000; 2003; 2007; 2010; 2011; 2015). Sponge also represent a key taxon, which has been widely investigated in terms of symbionts (Regoli *et al.*, 2004) and associated microbes (Lo Giudice *et al.*, 2019). In recent years also species new to science have been described, including the parasitic amphipod Lepidepecreella debroyeri (Schiaparelli *et al.*, 2004) and Adamussium colbecki (Schiaparelli and Linse, 2006). About the latter species, new analyses of data collected in 2006-2007, thanks to the presence of a mooring within the ASPA boundaries (Mooring "L" under the Italian mooring code system), showed that this species recruits during summer months in coincidence with an increase of the seawater temperature and a seasonal shift in the water currents and intensity (Schiaparelli and Aliani, 2019).

Below 70–75 m down to 120–130 m depth, heterogeneous substrates allow hard- and soft-bottom communities to coexist. On the sparse rocky outcrops the encrusting algae disappear and the benthic communities are dominated by the sessile zoobenthos. This diversified filter feeding assemblage is mainly characterised by sponges and anthozoans, while in soft sediments detritus-feeder polychaetes and bivalves dominate. Among sponges, which can reach very high biomass, Axociella nidificata, Calyx arcuarius, Gellius rudis, Phorbas glaberrima, Tedania charcoti, are very abundant (Sarà *et al.*, 1992; 2002; Gaino *et al.*, 1994; Cattaneo-Vietti *et al.*, 1996; 2000c; Bavestrello *et al.*, 2000; Cerrano *et al.*, 2000a). Numerous invertebrates constitute an important component of this assemblage which develops down to 120–140 m depth. These include crustacean peracarids, pycnogonids, mollusc opisthobranchs (Austrodoris kerguelenensis, Tritoniella belli) (Cattaneo-Vietti, 1991; Gavagnin *et al.*, 1995) and bivalves, ophiuroids and holothuroids, bryozoans, and a variety of endobionts. The conspicuous sponge spicule mats found at these depths underline the important role of sponges in this area, besides the one played by diatoms, in determining the sediment texture and silica content. A peculiar community, dominated by polychaetes and by the bivalve Limatula hodgsoni, can be associated with these mats.

Below 130 m the hard substrates become very sparse and are mainly colonised by the polychaete Serpula narconensis (Schiaparelli *et al.*, 2000) and several bryozoans (Arachnopusia decipiens, Ellisina antarctica, Flustra angusta, F. vulgaris and Isoschizoporella similis). The dominant muddy bottoms are instead characterised by tubicolous polychaetes (Gambi *et al.*, 2000b), mainly Spiophanes. Much deeper, at about 150-200 m depth, brachiopods and various species of bivalves characterise the environment on small gravels as well as on the soft bottom (Cattaneo-Vietti *et al.*, 2000b). The great heterogeneity of these substrates contributes to the creation of communities with considerable species richness, diversity and biomass. New polychaete species have also been described in 2021at ~ 500m of depth, i.e. the ampharetidAmphicteis teresae (Schiaparelli and Jirkov, 2016) and Amage giacomobovei (Schiaparelli & Jirkov, 2021).

#### Bird, fish and mammals

An Adélie penguin (Pygoscelis adeliae) colony is situated nearby the Area at Adélie Cove, with a 2013 population of 13,408 breeding pairs (Humphries *et al.*, 2017) (Map 1). About 30 Skua (Stercorarius maccormicki) pairs breed close to the penguins.

The faunal assemblage of the Area includes notothenioid fishes, represented especially by species of the Trematomus group, including T. bernacchi, T. pennelli, T. hansoni and T. loennbergii. These exert an important role in benthic food webs as consumers of many invertebrate species, mainly crustaceans and polychaetes (Vacchi *et al.*, 1991; 1992; 1994a; 1994b; 1995; 1997; 2000b; La Mesa *et al.*, 1996; 1997; 2000; Guglielmo *et al.*, 1998). The platelet ice occurring at Terra Nova Bay in early spring has been shown to house an important nursery for the Antarctic silverfish, Pleuragramma antarcticum, a key organism in the ecology of Antarctic food webs (La Mesa *et al.*, 2004; Vacchi *et al.*, 2004). The distribution of Pleuragramma eggs has been studied in detail only at three Terra Nova Bay sampling locations (i.e. Gerlache Inlet, Silverfish Bay and Cape Washington) (Guidetti *et al.*, 2015) where it was showing a certain degree of patchiness in eggs and larvae distribution (significantly changing at a spatial scale of kilometres and showing a not homogeneous distribution under the solid ice). No specific studies to quantify Pleuragramma eggs have been performed within the ASPA 161 boundaries, but it is know that abundant platelet ice is also occurring here in this area where additional nurseries of this fish species could reasonably be found. The platelet ice environment has also strong prooxidant characteristics at the beginning of austral spring, and the marked responsiveness of antioxidant defences represents a fundamental strategy for P. antarcticum (Regoli *et al.*, 2005b).

An aerial survey on cetacean species, conducted in the coastal area surrounding the Italian Station Mario Zucchelli, showed the presence of Killer Whale Orcinus orca (L.), types B and C and Minke Whale (Balaenoptera bonaerensis Burmeister). (Lauriano *et al.*, 2007a; 2007b.). Leopard seals (Hydrurga leptonix) were sighted several times at the end of the slope that penguins climb to reach the colony in the area represented in Map 1.



#### **Environmental characterization**

Studies on industrial pollutants in biomarkers allowed monitoring of the impact of human activities on the Antarctic biota in Terra Nova Bay area (Focardi *et al.*, 1995; Regoli *et al.*, 1998; Jimenez *et al.*, 1999; Regoli *et al.*, 2005a; Benedetti *et al.*, 2005, 2007; Canapa *et al.*, 2007; Di Bello *et al.*, 2007, Corsolini, 2009).

In Terra Nova Bay, organisms are exposed to a naturally elevated bioavailability of cadmium causing tissue concentrations generally 10-50 folds higher than those typical of temperate species (Mauri *et al.*, 1990; Nigro *et al.*, 1992, 1997; Canapa *et al.*, 2007, Caruso *et al.*, 2018). Elevated level of cadmium at Terra Nova Bay modulates bioaccumulation and metabolism of polycyclic aromatic hydrocarbons and of organochlorine xenobiotics in local marine organisms (Regoli *et al.*, 2005a; Benedetti *et al.*, 2007; Canapa *et al.*, 2007). Recent analyses (Signa *et al.* 2019) reported increased concentrations of Pb and Hg (Pb: Grotti *et al.*, 2008; lanni *et al.*, 2010; Hg: Bargagli *et al.*, 1998; Negri *et al.*, 2006), and phytoplankton reached trace elements levels from 2-fold (Hg) to 4-fold (Cd) and even 10-fold (Pb) higher than those previously recorded (Bargagli *et al.*, 1998; Dalla Riva *et al.*, 2004). In contrast, Hg concentration measured in feathers of Adelie penguins (Pygoscelies adelie) and Skua (Catharacta maccormlcki) in 2013 (Signa *et al.* 2019) did not differ from those measured in 1989-1991 (Bargagli *et al.* 1998).

A systematic publication of faunal check-lists for the Terra Nova Bay area has been started by the Italian National Antarctic Museum (MNA, https://steu.shinyapps.io/MNA-generale/) in 2013, with the final target to provide to GBIF distributional information for all taxa occurring in the area. Data are available for: Mollusca (Ghiglione *et al.*, 2013), Tanaidacea (Piazza *et al.*, 2014), Ophiuroidea (Cecchetto *et al.*, 2017), Porifera (Ghiglione *et al.*, 2018), Asteroidea (Moreau *et al.*, 2018; Guzzi *et al.*, 2022), Bryozoa (Cecchetto *et al.*, 2019), Rotifera (Garlaschè *et al.*, 2020), planktonic Copepoda (Bonello *et al.*, 2020), planktonic, benthic and sympagic copepods (Grillo *et al.*, 2024), Gnathostomulida (Sterrer *et al.*, 2022), polynoid polychaetes (Cowart et asl., 2022), fishes (La Mesa *et al.*, 2022), echinoids and crinoids (Guzzi *et al.*, 2023). ).

Long-term monitoring activities using non-destructive sampling techniques such as ROV, benthic cameras and ROV surveys have also been carried out in the Terra Nova Bay in recent years, further defining the set of organisms occurring in the area (Canese *et al.*, 2015; Piazza *et al.*, 2018; 2019; 2020; Castellan *et al.*, 2021; La Mesa *et al.*, 2022; Marini *et al.*, 2022a; 2022b).

Ongoing studies on food web structure will enable quantification of trophic interactions between species and potential community vulnerability to biodiversity loss and changes in sea-ice dynamics (Calizza *et al.*, 2018, Signa *et al.*, 2019; Rossi *et al.*, 2019).

#### **Human Activities**

The Area is close to the Italian Station Mario Zucchelli (74°41'39"S,164°06'55"E) that can accommodate approximately 90 people, has facilities for helicopter operations and a jetty for the docking of small boats. Fuel used at the station is Jet-A1. The station is equipped with a waste water treatment plant. Treated water is discharged into the sea adjacent to the station 2.3 km from the northern boundary of the Area. A support ship regularly visits Mario Zucchelli Station during the summer, and there are occasional visits by tourist ships. These usually stop offshore several kilometers to the north of the Area.

Other nearby stations are Gondwana (74°38'0.7"S, 164°13'19" E; Germany), a summer station with capacity for approximately 25 personnel, Jang Bogo station (74°37'15"S ,164°11'57"E; Republic of Korea) year round station with a complement of 60 personnel during summer and 17 during winter, the new Qinling station (74°56'0.4"S, 163°42'55"E; China) at Inexpressible Island, a year-round station with a complement of up to 30 in winter and 80 summer personnel.

A gravel runway has been built at Boulder Clay site, Terra Nova Bay (74°44′45″S, 164°01′17″E, 205 m a.s.l.). The end of the runway is about 1.8 km from the penguin colony of Adelie Cove. An Environmental Impact Monitoring Plan has been developed to evaluate changes in the ecosystem during construction and operation of the runway (Draft CEE – MZS gravel runway ATCM39).

#### 6 (ii) Access to the Area

Access into the Area is generally by ship. Access into the Area may be made by air or over sea ice when conditions allow. Access routes within the Area have not been defined.

#### 6 (iii) Location of structures within and adjacent to the Area

There are no permanent structures within the Area. The nearest structure is the atmospheric monitoring facility (locally referred to as 'Campo Icaro') 650 m north of the northern boundary of the Area, while Mario Zucchelli Station (74°41'42"S, 164°07'23"E) is situated on a small peninsula on the coast adjacent to Tethys Bay, a further 1.65 km to the north. A gravel runway is located at Boulder Clay site, Terra Nova Bay (74°44'45"S, 164°01'17"E, 205 m a.s.l.). The end of the runway is about 1.8 km from the penguin colony of Adelie Cove. The Italian Marine Observatory in the Ross Sea (MORSea) maintains a mooring within the Area (74°45'25"S, 163°42'55"E).

#### 6 (iv) Location of other protected areas in the vicinity

ASPA No. 178 Inexpressible Island and Seaview Bay is situated about 17 km to the south; ASPA No. 175 the high altitude geothermal sites on Mount Melbourne, is a terrestrial site situated 45 km to the NE.

#### 6 (v) Special zones within the Area

There are no special zones within the Area.



# 7. Terms and conditions for entry permits

## 7 (i) General permit conditions

Entry into the Area is prohibited except in accordance with a permit issued by the appropriate national authority. Conditions for issuing a permit are that:

- it is issued for scientific purposes, or for educational purposes which cannot be served elsewhere;
- it is issued for essential management purposes consistent with plan objectives such as inspection, maintenance or review;
- the actions permitted will not jeopardise the values of the Area;
- any management activities are in support of the objectives of the Management Plan;
- the actions permitted are in accordance with the Management Plan;
- the permit, or a copy, shall be carried by the holder within the Area;
- permits shall be issued for a stated period.

## 7 (ii) Access to and movement within the Area

Access into the Area shall be by sea, land, over sea ice or by air. There are no specific restrictions on routes of access to and movement within the Area, although movements should be kept to the minimum necessary consistent with the objectives of any permitted activities and every reasonable effort should be made to minimise disturbance. Anchoring is prohibited within the Area. There are no overflight restrictions within the Area and aircraft may land by permit when sea ice conditions allow, taking into consideration the penguin colony situated at Adelie Cove and following the Guidelines for Operations of Aircraft near Concentration of Birds in Antarctica (Resolution 2, 2004), to limit disturbance.

Overflight and landings within the Area by Remotely Piloted Aircraft Systems (RPAS) are prohibited except in accordance with a permit issued by an appropriate national authority. RPAS use within the Area should follow the Environmental Guidelines for Operation of Remotely Piloted Aircraft Systems (RPAS) in Antarctica (Resolution 4 (2018).

## 7 (iii) Activities that may be conducted within the Area

Activities that may be conducted in the Area should not jeopardise the values of the Area and include:

- Scientific research that cannot be served elsewhere;
- Sampling, which should be the minimum required to reach the scientific goals. Selective and less-invasive sampling methods should always be considered to reduce disturbance of the rich bottom communities;
- Essential management activities, including monitoring and inspection;
- Operational activities in support of scientific research or management of the Area;
- Activities for educational and outreach purposes.

#### 7 (iv) Installation, modification or removal of structures

Structures or scientific equipment shall not be installed within the Area except as specified in a permit. All markers, structures or scientific equipment installed in the Area shall be clearly identified by country, name of the principal investigator and year of installation. All such items should be made of materials that pose minimal risk of contamination of the Area. Removal of specific equipment for which the permit has expired is mandatory.

## 7 (v) Location of field camps

None within the Area.

## 7 (vi) Restrictions on materials and organisms which can be brought into the Area

- No living animals, plant material, pathogens or microorganisms shall be deliberately introduced into the Area.
- Poultry products, including food products containing uncooked dried eggs, shall not be introduced into the Area.
- No herbicides or pesticides shall be introduced into the Area.
- Chemicals, including radio-nuclides or stable isotopes, which may be introduced for the scientific or management purposes specified in the permit, shall be used in the minimum quantities necessary to achieve the purpose of the activity for which the permit was granted.
- All materials introduced in the Area shall be stored and handled so that risk of their accidental release into the environment is minimized and removed at the end of the period allowed in the permit.
- Visitors shall take special precautions against marine pollution and ensure that sampling equipment or markers brought into the Area are clean. Vessels that are found to show fuel leakage, or a significant risk of such leakage, are prohibited from entering the Area.



## 7 (vii) Taking or harmful interference with native flora or fauna

Taking or harmful interference with native flora or fauna is prohibited, except by permit issued in accordance with Annex II to the Protocol on Environmental Protection to the Antarctic Treaty. Careful environmental evaluation is needed concerning trawling, dragging, grabbing, dredging, or deployment of nets because of the sensitivity of the rich bottom communities to disturbance. More selective and less-invasive sampling methods should always be considered;

Where taking of or harmful interference with animals is involved, the SCAR Code of Conduct for the Use of Animals for Scientific Purposes in Antarctica (ATCM XXXIV-CEP XIV, 2011) should be used as a minimum standard.

#### 7 (viii) Collection or removal of materials not brought into the Area by the permit holder

Any anthropogenic material found should be notified to the appropriate national authority.

Material may be collected or removed from the Area only in accordance with a permit. In this case removal of material should not create an impact greater than leaving the material in situ.

## 7 (ix) Disposal of waste

All wastes, including all human wastes, shall be removed from the Area.

#### 7 (x) Measures that may be necessary to continue to meet the aims of the Management Plan

Permits may be granted to enter the Area to

- carry out monitoring and site inspection activities, which may involve the collection of limited samples for analysis or review, or for protective measures;
- Install markers on specific sites of long-term monitoring.

### 7 (xi) Requirements for reports

The holder of each permit issued should report to the appropriate national competent authority about the activity undertaken in the Area.

Such reports should include, as appropriate, the information identified in Appendix 2- ASPA visit report form of the Guide to the Preparation of Management plans for ASPAs (Resolution 2, 2011). Parties should, wherever possible, exchange with the Party that proposed the Management Plan, information on reports received to assist managing the Area.

## 8. References and Relevant Supporting Bibliography

Accornero A., Manno C., Arrigo K.R., Martini Atucci S., 2003. "The vertical flux of particulate matter in the polynya of Terra Nova Bay. Part I. Chemical constituents" Antarctic Science 15 (1), 119–132.

Ademollo N., Cincinelli A., Sarà G., Vecchiato M., Corsolini S., 2022. Influence of climate change on bioaccumulation pattern of legacy and emerging Persistent organic pollutants in Antarctica, In SCAR 2022: Antarctica in a Changing World, SCAR Open Science Conference 2022 - Full Abstract Book. 10th SCAR Open Science Conference. Pag 474.

Ademollo N., Pala N., Baroni D., Corsolini S., 2021. Temporal Trend of Current-Use Pesticides in Adèlie Penguin Eggs From the Ross Sea (Antarctica). Society of Environmental Toxicology and Chemistry North America 42nd Annual Meeting – SETAC Solutions With Respect for Our Community and Environment, 14–18 November 2021

Ademollo N., Corsolini S., Spataro F., Rauseo J., Pescatore T., Valsecchi S., Polesello S., Patrolecco L., 2019. Temporal trend of perfluoroalkyl substances and current-use pesticides in penguin eggs from the Ross Sea (Antarctica). Abstract Book, Society of Environmental Toxicology and Chemistry North America 40th Annual Meeting, Great Together: Separate Challenges and Collective Solutions 3–7 November 2019, Toronto, Ontario, Canada. Pp. 339.

Albertelli G., Cattaneo-Vietti R., Chiantore M., Pusceddu A., Fabiano M., 1998. Food availability to an *Adamussium* bed during the austral Summer 1993/94 (Terra Nova Bay, Ross Sea). Journal of Marine Systems 17: 425-34.

Alongi, G., Cormaci, M. & Furnari, G. (2002). The Corallinaceae (Rhodophyta) from the Ross Sea (Antarctica): a taxonomic revision rejects all records except *Phymatolithon foecundum*. Phycologia, 41: 140–146.

Alvaro M.C, Blazewicz-Paszkowycz M., Davey N., Schiaparelli S., 2011. Skin-digging tanaidaceans: the unusual parasitic behaviour of *Exspina typica* (Lang, 1968) in Antarctic waters and worldwide deep basins. Antarct Sci, vol. 23 (4); p. 343-348.

Alvizu, A., Xavier, J. R., & Rapp, H. T. (2019). Description of new chiactine-bearing sponges provides insights into the higher classification of Calcaronea (Porifera: Calcarea). Zootaxa, 4615(2), 201-251.

Ansell A.D., Cattaneo-Vietti R., Chiantore M., 1998. Swimming in the Antarctic scallop *Adamussium colbecki*: analysis of in situ video recordings. Antarctic Science 10 (4): 369-75.

Ballerini T., Tavecchia G., Olmastroni S., Pezzo F., Focardi S., 2009. Nonlinear effects of winter sea ice on the survival probabilities of Adélie penguins. Oecologia 161:253–265.

Bargagli R., Nelli L., Ancora S., Focardi S., 1996. Elevated cadmium accumulation in marine organisms from Terra Nova bay (Antartica). Polar Biology 16: 513-520.



Bargagli R., Monaci F., Sanchez-Hernandez J.C., Cateni D., 1998. Biomagnification of mercury in an Antarctic marine coastal food web. Marine Ecology Progress Series 169: 65-76.

Bargagli R.,2005. Antarctic Ecosystems. Environmental Contamination, Climate Change, and Human Impact. Ecological Studies, vol. 175; Springer-Verlag, Heidelberg, 395 pp.

Bargagli R., 2008. Environmental contamination in Antarctic ecosystems. Sci. Total Environ. 400: 212-226.

Bargelloni, L., Babbucci, M., Ferraresso, S., Papetti, C., Vitulo, N., Carraro, R., Pauletto, M., Santovito, G., Lucassen, M., Mark, F.C., Zane, L., Patarnello, T (2019). Draft genome assembly and transcriptome data of the icefish *Chionodraco myersi* reveal the key role of mitochondria for a life without hemoglobin at subzero temperatures. Communications Biology, 2 (1), art. no. 443. DOI: 10.1038/s42003-019-0685-y

Bavestrello G., Arillo A., Calcinai B., Cattaneo-Vietti R., Cerrano C., Gaino E., Penna A., Sara' M., 2000. Parasitic diatoms inside Antarctic sponges. Biol. Bull. 198: 29-33.

Benedetti M., Gorbi S., Bocchetti R., Fattorini D., Notti A., Martuccio G., Nigro M., Regoli F. (2005). Characterization of cytochrome P450 in the Antarctic key sentinel species *Trematomus bernacchii*. Pharmacologyonline 3: 1-8 ISSN-1827-8620.

Benedetti M., Martuccio G., Fattorini D., Canapa A., Barucca M., Nigro M., Regoli F. (2007). Oxidative and modulatory effects of trace metals on metabolism of polycyclic aromatic hydrocarbons in the Antarctic fish *Trematomus bernacchii*. Aquat. Toxicol. 85: 167-175

Berkman P.A., Nigro M., 1992. Trace metal concentrations in scallops around Antarctica: Extending the Mussel Watch Programme to the Southern Ocean. Marine Pollution Bulletin 24 (124): 322-23.

Bonello G., Grillo M., Cecchetto M., Giallain M., Granata A., Guglielmo L., Pane L., Schiaparelli S. (2020) Distributional records of Ross Sea (Antarctica) planktonic Copepoda from bibliographic data and samples curated at the Italian National Antarctic Museum (MNA): checklist of species collected in the Terra Nova Bay area (western Ross Sea) from 1987 to 1995. Zookeys, 969: 1–22.

Borghesi N., Corsolini S., Focardi S., 2008. Levels of polybrominated diphenyl ethers (PBDEs) and organochlorine pollutants in two species of Antarctic fish (*Chionodraco hamatus* and *Trematomus bernacchii*). Chemosphere, 73, 155–160.

Bruni V., Maugeri M.L., Monticelli L.S., 1997. Faecal pollution indicators in the Terra Nova Bay (Ross Sea, Antarctica). Marine Pollution Bulletin 34 (11): 908–12.

Budillon G., Spezie G., 2000. "Thermoaline structure and variability in the Terra Nova Bay polynya (Ross Sea) between 1995-98". Antarctic science 12, 243-254.

Calizza E., Careddu G., Sporta Caputi S., Rossi L., Costantini M.L., 2018. Time-and depth-wise trophic niche shifts in Antarctic benthos. PloS one 13: e0194796.

Canapa A., Barucca M., Gorbi S., Benedetti M., Zucchi S., Biscotti MA., Olmo E., Nigro M., Regoli F., 2007. Vitellogenin gene expression in males of the Antarctic fish *Trematomus bernacchii* from Terra Nova Bay (Ross Sea): A role for environmental cadmium? Chemosphere, 66:1270–1277.

Canese S., Mazzoli C., Montagna P., Schiaparelli S., Taviani M., 2015. The Terra Nova Bay 'Canyon': ROV survey of nearshore shallow to deep carbonate factories. XII International Symposium on Antarctic Earth Sciences ISAES, 13-17 July 2015, Goa, India.

Cantone G., Castelli A., Gambi M.C., 2000. The Polychaete fauna off Terra Nova Bay and Ross Sea: biogeography, structural aspects and ecological role. In: Ross Sea Ecology, F. Faranda, L. Guglielmo and A. Ianora Eds., Springer Verlag, Berlin Heidelberg: 551-61.

Caruso C., Rizzo C., Mangano S., Poli A., Di Donato P., Nicolaus B., Di Marco G., Michaud L., Lo Giudice A. (2018). Extracellular polymeric substances with metal adsorption capacity produced by *Pseudoalteromonas* sp. MER144 from Antarctic seawater. Environmental Science and Pollution Research, 25: 4667-4677.

Castellan G., Angeletti L., Canese S., Mazzoli C., Montagna P., Schiaparelli S., Taviani M. (2021) Visual Imaging of Benthic Carbonate–Mixed Factories in the Ross Sea Region Marine Protected Area, Antarctica. Minerals, 11, 833.

Castellano M. 2006. "Aspetti trofo-funzionali dell'ecosistema marino costiero antartico: sostanza organica particellata e disciolta", Univeristà degli Studi di Genova, PhD Thesys.

Cattaneo-Vietti R., 1991. Nudibranch Molluscs from the Ross Sea, Antarctica. J. Moll. Stud. 57: 223-28.

Cattaneo-Vietti R., Bavestrello G., Cerrano C., Gaino E., Mazzella L., Pansini M., Sarà M., 2000c. The role of sponges of Terra Nova Bay ecosystem. In: Ross Sea Ecology, F. Faranda, L. Guglielmo and A. Ianora Eds., Springer Verlag, Berlin Heidelberg: 539-49.

Cattaneo-Vietti R., Bavestrello G., Cerrano C., Sara' M., Benatti U., Giovine M., Gaino E., 1996. Optical fibres in an Antarctic sponge. Nature 383: 397-98.

Cattaneo-Vietti R., Chiantore M., Albertelli G., 1997. The population structure and ecology of the Antarctic Scallop, *Adamussium colbecki* in Terra Nova Bay (Ross Sea, Antarctica). Scientia Marina 61 (Suppl. 2): 15-24.

Cattaneo-Vietti R., Chiantore M., Gambi M.C., Albertelli G., Cormaci M., Di Geronimo I., 2000a. Spatial and vertical distribution of benthic littoral communities in Terra Nova Bay. In: Ross Sea Ecology, F. Faranda, L. Guglielmo and A. Ianora Eds., Springer Verlag, Berlin Heidelberg: 503-14.

- (



Cattaneo-Vietti R., Chiantore M., Misic C., Povero P., Fabiano M., 1999. The role of pelagic-benthic coupling in structuring littoral benthic communities at Terra Nova Bay (Ross Sea) and inside the Strait of Magellan. Scientia Marina 63 (Supl. 1): 113-21.

Cattaneo-Vietti R., Chiantore M., Schiaparelli S., Albertelli G., 2000b. Shallow and deep-water mollusc distribution at Terra Nova Bay (Ross Sea, Antarctica). Polar Biology 23: 173-82.

Cecchetto M., Alvaro M.C., Ghiglione C., Guzzi A., Mazzoli C., Piazza P., Schiaparelli S., 2017. Distributional records of Antarctic and sub-Antarctic Ophiuroidea from samples curated at the Italian National Antarctic Museum (MNA): check-list update of the group in the Terra Nova Bay area (Ross Sea) and launch of the MNA 3D model 'virtual gallery'. ZooKeys, 705: 61-79.

Cecchetto M., Lombardi C., Canese S., Cocito S., Kuklinski P., Mazzoli C., Schiaparelli S., 2019. Bryozoa collection of the Italian National Antarctic Museum (MNA), with an updated checklist from Terra Nova Bay (Ross Sea). Zookeys 812: 1-22.

Cecchetto M., Di Cesare A., Eckert E., Fassio G., Fontaneto D., Moro I., Oliverio M., Sciuto K., Tassistro G., Vezzulli L., Schiaparelli S. (2021) Antarctic coastal nanoplankton dynamics revealed by metabarcoding of desalination plant filters: Detection of short-term events and implications for routine monitoring. Science of the Total Environment, 757 (2021) 143809.

Cerrano C., Calcinai B., Cucchiari E., Di Camillo C., Nigro M., Regoli F., Sarà A., Schiaparelli S., Totti C., Bavestrello G., 2004. Are diatoms a food source for Antarctic sponges?. Chemistry and Ecology, vol. 20: 57-64.

Cerrano C., Arillo A., Bavestrello G., Calcinai B., Cattaneo-Vietti R., Penna A., Sarà M., Totti C., 2000a. Diatom invasion in the Antarctic hexactinellid sponge *Scolymastra joubini*. Polar Biology 23: 441-44.

Cerrano C., Bavestrello G., Calcinai B., Cattaneo-Vietti R., Sarà A., 2000b. Asteroids eating sponges from Tethys Bay, East Antarctica. Antarctic Science 12(4): 431-32.

Cerrano C., G. Bavestrello, B. Calcinai, R. Cattaneo-Vietti, M. Chiantore, M. Guidetti, A. Sarà, 2001a. Bioerosive processes in Antarctic seas. Polar Biology 24: 790–92.

Cerrano C., Puce S., Chiantore M., Bavestrello G., 2000c. Unusual trophic strategies of *Hydractinia angusta* (Cnidaria, Hydrozoa) from Terra Nova Bay, Antarctica. Polar Biology 23(7): 488-94.

Cerrano C., S. Puce, M. Chiantore, G. Bavestrello, R. Cattaneo-Vietti, 2001b. The influence of the epizooic hydroid *Hydractinia angusta* on the recruitment of the Antarctic scallop *Adamussium colbecki*. Polar Biology 24: 577-81.

Chatzidimitriou E., Bisaccia P., Corrà F., Bonato M., Irato P., Manuto L., Toppo S., Bakiu R., Santovito G. (2020) Copper/zinc superoxide dismutase from the crocodile icefish *Chionodraco hamatus*: antioxidant defense at constant sub-zero temperature. Antioxidants 9, 325.

Chiantore M., Cattaneo-Vietti R., Albertelli G., Misic M., Fabiano M., 1998. Role of filtering and biodeposition by *Adamussium colbecki* in circulation of organic matter in Terra Nova Bay (Ross Sea, Antarctica). Journal of Marine Systems 17: 411-24.

Chiantore M., Cattaneo-Vietti R., Berkman P.A., Nigro M., Vacchi M., Schiaparelli S., Albertelli G., 2001. Antarctic scallop (*Adamussium colbecki*) spatial population variability along the Victoria Land Coast, Antarctica. Polar Biology 24: 139-43.

Chiantore M., Cattaneo-Vietti R., Povero P., Albertelli G., 2000. The population structure and ecology of the antarctic scallop *Adamussium colbecki* in Terra Nova Bay. In: Ross Sea Ecology, F. Faranda, L. Guglielmo and A. Ianora Eds., Springer Verlag, Berlin Heidelberg: 563-73.

Chiantore M., R. Cattaneo-Vietti, L. Elia, M. Guidetti, M. Antonini, 2002. Reproduction and condition of the scallop *Adamussium colbecki* (Smith 1902), the sea-urchin *Sterechinus neumayeri* (Meissner, 1900) and the sea-star Odontaster validus Koehler, 1911 at Terra Nova Bay (Ross Sea): different strategies related to inter-annual variations in food availability. Polar Biology 22: 251-55.

Chiantore M., Vacchi M., 2012. Dense populations of the Antarctic scallop *Adamussium colbecki* in Terra Nova Bay (Subarea 88.1J): potential VMEs adiacent to the Terra Nova Bay ASPA (No. 161). CCAMLR WG-MME-12/23, 12 pp.

Clark M.S., Hoffman J.I., Peck L.S., *et al.*, (2023). Multi-omics for studying and understanding polar life. Nature Communications, 14, 1, D0I10.1038/s41467-023-43209-y

Cormaci M., Furnari G., Scammacca B., 1992b. The benthic algal flora of Terra Nova Bay (Ross Sea, Antarctica). Botanica Marina 35(6): 541-52

Cormaci M., Furnari G., Scammacca B., 1992c. Carta della vegetazione marina di Baia Terra Nova (Mare di Ross, Antartide). Biologia Marina 1: 313–14.

Cormaci M., Furnari G., Scammacca B., Alongi G., 1996. Summer biomass of a population of *Iridaea cordata* (Gigartinaceae, Rhodophyta) from Antarctica. In: Lindstrom SC, Chapman DJ (Eds) Proceedings of the XV Seeweeds Symposium. Hydrobiologia 326/327: 267-72.

Cormaci M., Furnari G., Scammacca B., Casazza G., 1992a. Il fitobenthos di Baia Terra Nova (Mare di Ross, Antartide): osservazioni sulla flora e sulla zonazione dei popolamenti. In: Gallardo VA, Ferretti O, Moyano HI (eds) Actas del Semin. Int. Oceanografia in Antartide. Centro EULA, Universitad de Concepción, Chile. ENEA: 395-408.

Corsolini S, Nigro M, Olmastroni S, Focardi S, Regoli F 2001 Susceptibility to oxidative stress in Adelie and Emperor penguin, Polar Biology, vol. 24: 365–368.

Corsolini S. Borghesi N., Ademolo N., Focardi S., 2011. Chlorinated biphenyls and pesticides in migrating and resident seabirds from East and West Antarctica. Environment International 37(8): 1329-1335.



Corsolini S., 2009. Industrial contaminants in Antarctic biota. Journal of Chromatography A, 1216, 598-612.

Corsolini S., 2011. Antarctic: Persistent Organic Pollutants and Environmental Health in the Region. In: Nriagu JO (ed.) Encyclopedia of Environmental Health, volume 1, pp. 83–96 Burlington: Elsevier,NVRN/978-0-444-52273-3.

Corsolini S., Kannan K., Imagawa T., Focardi S., Giesy J.P., 2002. Polychloronaphthalenes and other dioxin-like compounds in Arctic and Antarctic marine food webs. Environmental Science and Technology, 36: 3490-3496.

Corsolini S., Ademollo N., 2022. POPs in Antarctic ecosystems: is climate change affecting their temporal trends? Environ. Sci.: Processes Impacts, 24(10), 1631-1642, https://doi.org/10.1039/D2EM00273F

Corsolini S., Ancora S., Ademollo N., Jiménez B, 2022. Editorial: From Pole to Pole: Contamination of Marine Ecosystems in a Changing World. Front. Mar. Sci. 9:899494. doi:10.3389/fmars.2022.899494.

Cowart D.A., Schiaparelli S., Alvaro M.C., Cecchetto M., Le Port A.-S., Jollivet D., Hourdez S. (2022) Origin, diversity, and biogeography of Antarctic scale worms (Polychaeta: Polynoidae): a wide-scale barcoding approach. Ecology and Evolution: 12:e9093.

Dalla Riva S., Abelmoschi M.L., Magi E., Soggia F., 2004. The utilization of the antarctic environmental specimen bank (BCAA) in monitoring Cd and Hg in an antarctic coastal area in Terra Nova bay (Ross Sea - Northern Victoria land). Chemosphere 56: 59-69.

Di Bello D., Vaccaio E., Longo V., Regoli F., Nigro M., Benedetti M., Gervasi PG, Pretti C. (2007). Presence and inducibility by ß-Naphtoflavone of CYP 1A1, CYP 1B1, UDP-GT, GST and DT-Diaphorase enzymes in *Trematomus bernacchii*, an Antarctic fish. Aquatic Toxicol. 84: 19-26

Fabiano M., Chiantore M., Povero P., Cattaneo-Vietti R., Pusceddu A., Misic C., Albertelli G., 1997. Short-term variations in particulate matter flux in Terra Nova Bay, Ross Sea. Antarctic Science 9(2): 143-149.

Fabiano M., Danovaro R., Crisafi E., La Ferla R., Povero P., Acosta Pomar L., 1995. Particulate matter composition and bacterial distribution in Terra Nova Bay (Antarctica) during summer 1989-90. Polar Biology 15: 393-400.

Fabiano M., Povero P., Danovaro R., 1996. Particulate organic matter composition in Terra Nova Bay (Ross Sea, Antarctica) during summer 1990. Antarctic Science 8(1): 7-13.

Fassio G., Buge B., Salvi D., Oliverio M., Alvaro M.C., Modica M.V., Schiaparelli S. (2019) An Antarctic flock under the Thorson's rule: diversity and larval development of Antarctic Velutinidae (Mollusca: Gastropoda). Molecular Phylogenetics and Evolution, 132: 1-13.

Focardi S., Fossi M.C., Lari L., Casini S., Leonzio C., Meidel S.K., Nigro M., 1995. Induction of MFO Activity in the Antarctic fish *Pagothenia bernacchii*: Preliminary results. Marine Environmental Research., 39: 97-100.

Focardi S., Bargagli R., Corsolini S., 1993. Organochlorines in marine Antarctic food chain at Terra Nova Bay (Ross Sea). Korean Journal of Polar Research 4: 73-77.

Frezzotti, M., Salvatore, M.C., Vittuari, L., Grigioni, P., De Silvestri L., 2001. Satellite Image Map: Northern Foothills and Inexpressible Island Area (Victoria Land, Antarctica). Terra Antarctica Reports n° 6, 8 p. + map – ISBN 88-900221-9-1.

Gaino E., Bavestrello G., Cattaneo-Vietti R., Sara' M., 1994. Scanning electron microscope evidence for diatom uptake by two Antarctic sponges. Polar Biology 14: 55-58.

Gambi M.C., Buia M.C., Mazzella L., Lorenti M., Scipione M.B., 2000a. Spatio-temporal variability in the structure of benthic populations in a physically controlled system off Terra Nova Bay: the shallow hard bottoms. In: Ross Sea Ecology, F. Faranda, L. Guglielmo and A. Ianora Eds., Springer Verlag, Berlin Heidelberg: 527-538.

Gambi M.C., Castelli A., Guizzardi M., 1997. Polychaete populations of the shallow soft bottoms off Terra Nova Bay (Ross Sea, Antarctica): distribution, diversity and biomass. Polar Biology 17: 199–210.

Gambi M.C., Giangrande A., Patti F.P., 2000b. Comparative observations on reproductive biology of four species of *Perkinsiana* (Polychaeta, Sabellidae). Bulletin of Marine Science 67(1): 299-309.

Gambi M.C., Lorenti M., Russo G.F., Scipione M.B., 1994. Benthic associations of the shallow hard bottoms off Terra Nova Bay (Ross Sea, Antarctica): zonation, biomass and population structure. Antarctic Science 6(4): 449-62.

Gavagnin M., Trivellone E., Castelluccio F., Cimino G., Cattaneo-Vietti R., 1995. Glyceryl ester of a new halimane diterpenoic acid from the skin of the antarctic nudibranch *Austrodoris kerguelenensis*. Tetrahedron Letters 36: 7319-22.

Ghiglione C, Alvaro M.C., Griffiths H.J., Linse K., Schiaparelli S., 2013. Ross Sea Mollusca from the Latitudinal Gradient Program: R/V Italica 2004 Rauschert dredge samples. ZooKeys, 341: 37-48.

Garlaschè G., K. Karimullah, N. lakovenko, A. Velasco-Castrillón, K. Janko, R. Guidetti, L. Rebecchi, M. Cecchetto, S. Schiaparelli, C. D. Jersabek, W. H. De Smet, D. Fontaneto (2020) A data set on the distribution of Rotifera in Antarctica. Biogeographia – The Journal of Integrative Biogeography ,35: 17-25.

Ghiglione C., Alvaro M.C., Cecchetto M., Canese S., Downey R., Guzzi A., Mazzoli C., Piazza P., Rapp H.T., Sarà A., Schiaparelli S. (2018) Distributional records of Antarctic Porifera from samples stored at the Italian National Antarctic Museum (MNA), with an update of the checklist for the Terra Nova Bay area (Ross Sea). Zookeys, 758: 137-156.

Guidetti, P., Ghigliotti, L., & Vacchi, M. (2015). Insights into spatial distribution patterns of early stages of the Antarctic silverfish, *Pleuragramma antarctica*, in the platelet ice of Terra Nova Bay, Antarctica. Polar Biology, 38, 333–342.



Grillo M., Huettmann F., Guglielmo L., Schiaparelli S. (2022) Three-Dimensional Quantification of Copepods Predictive Distributions in the Ross Sea: First Data Based on a Machine Learning Model Approach and Open Access (FAIR) Data. Diversity, 14, 355.

Grillo M., Bonello G., Cecchetto M., Guzzi A., Noli N., Cometti V., Schiaparelli S. (2024) Planktonic, benthic and sympagic copepods collected from the desalination unit of Mario Zucchelli Research Station in Terra Nova Bay (Ross Sea, Antarctica). Biodiversity Data Journal. doi: 10.3897/BDJ.12.e119633.

Grotti M., Soggia F., Lagomarsino C., Dalla Riva S., Goessler W., Francesconi, K.A., 2008. Natural variability and distribution of trace elements in marine organisms from Antarctic coastal environments. Antarctic Science 20: 39-51.

Guglielmo G., Zagami G., Saggiorno V., Catalano G., Granata A., 2007. "Copepods in spring annual sea ice at Terra Nova Bay (Ross Sea, Antarctica)" Polar Biology 30, 747-758.

Guglielmo L., Carrada G.C., Catalano G., Dell'Anno A., Fabiano M., Lazzara L., Mangoni O., Pusceddu A., Saggiomo V., 2000. Structural and functional properties of sympagic communities in the annual sea ice at Terra Nova Bay (Ross Sea, Antarctica). Polar Biology 23(2): 137-46.

Guglielmo L., Granata A., Greco S., 1998. Distribution and abundance of postlarval and juvenile *Pleuragramma antarticum* (Pisces, Nototheniidae) of Terra Nova Bay (Ross Sea, Antartica). Polar Biology 19: 37-51.

Guzzi A., Alvaro M.C., Danis B., Moreau C., Schiaparelli S. (2022) Not all that glitters is gold: barcoding effort reveals taxonomic incongruences in iconic Ross Sea seastars. Diversity, 14(6), 457.

Guzzi A., Alvaro M.C., Cecchetto M., Schiaparelli S. (2023) Echinoids and Crinoids from Terra Nova Bay (Ross Sea) Based on a Reverse Taxonomy Approach. Diversity, 15, 875.

Humphries G.R.W., Che-Castaldo C., Naveen R., Schwaller M., McDowall P., Schrimpf M., and Lynch H.J. 2017. Mapping Application for Penguin Populations and Projected Dynamics (MAPPPD): Data and tools for dynamic management and decision support. Polar Records .

Ianni C., Magi E., Soggia F., Rivaro P., Frache R., 2010. Trace metal speciation in coastal and off-shore sediments from Ross Sea (Antarctica). Microchemical Journal 96: 203-212

Jimenez B., Fossi M.C., Nigro M., Focardi S., 1999. Biomarker approach to evaluating the impact of scientific stations on the Antarctic environment using *Trematomus bernacchii* as a bioindicator organism. Chemosphere, 39: 2073-2078.

La Mesa M., Arneri E., Giannetti G., Greco S., Vacchi M., 1996. Age and growth of the nototheniid fish *Trematomus bernacchii* Boulenger from Terra Nova Bay, Antartica. Polar Biology16: 139-45.

La Mesa M., J.T. Eastman, M. Vacchi, 2004. The role of notothenioid fish in the food web of the Ross Sea shelf waters: a review. Polar Biol., 27: 321-338.

La Mesa M., Vacchi M., Castelli A., Diviacco G., 1997. Feeding ecology of two nototheniid fishes *Trematomus hansoni* and *Trematomus loennbergi* from Terra Nova Bay, Ross Sea. Polar Biology 17: 62-68.

La Mesa M., Vacchi M., T. Zunini Sertorio, 2000. Feeding plasticity of *Trematomus newnesi* (Pisces, Nototheniidae) in Terra Nova Bay, Ross Sea, in relation to environmental conditions. Polar Biology 23(1): 38-45.

La Mesa M., Canese S., Montagna P; Schiaparelli S. (2022) Underwater Photographic Survey of Coastal Fish Community of Terra Nova Bay, Ross Sea. Diversity, 14, 315. https://doi.org/10.3390/d14050315.

Lauriano G., Fortuna C.M., Vacchi M., 2007a. Observation of killer whale (Orcinus orca)

Lauriano G., Vacchi M., Ainley D., Ballard G., 2007b. Observations of top predators foraging on fish in the pack ice of the southern Ross Sea. Antarctic Science, 19(4): 439-440.

Lo Giudice A., Casella P., Bruni V., Michaud L. (2013). Response of bacterial isolates from Antarctic shallow sediments towards heavy metals, antibiotics and polychlorinated biphenyls. Ecotoxicology, 22: 240-250

Lo Giudice A., Azzaro M., Schiaparelli S. (2019). Microbial Symbionts of Antarctic Marine Benthic Invertebrates. In The Ecological Role of Micro-organisms in the Antarctic Environment, Castro-Sowinski S. (Ed.), Chapter 13, Springer Polar Sciences. Pp. 277-296. https://doi.org/10.1007/978-3-030-02786-5\_13.

Mangoni O., Modigh M., Conversano F., Carrada G.C., Saggiorno V., 2004. "Effects of summer ice coverege on phytoplankton assemblages in the Ross Sea, Antarctica" Deep-Sea Research I, 51, 1601–1617.

Marini S., Federico B., Lorenzo C., Bordone A., Schiaparelli S., Peirano A. (2022a) Long-term Automated Visual Monitoring of Antarctic Benthic Fauna. Methods in Ecology and Evolution 13, 1746–1764.

Marini S., Bonofiglio F., Corgnati L.P., Bordone A., Schiaparelli S., Peirano A. (2022b) Long-term High Resolution Image Dataset of Antarctic Coastal Benthic Fauna. Scientific Data, 9:750.

Massolo S., Messa R., Rivaro P., Leardi R., 2009. "Annual and spatial variations of chemical and physical properties in the Ross Sea surface waters (Antarctica)" Continental Shel Research 29, 2333–2344.

Mauri M., Orlando E., Nigro M., Regoli F., 1990. Heavy metals in the Antarctic scallop *Adamussium colbecki* (Smith). Mar. Ecol. Progr. Ser. 67: 27-33.



Minganti V., Capelli R., Fiorentino F., De Pellegrini R., Vacchi M., 1995. Variations of mercury and selenium concentrations in *Adamussium colbecki* and *Pagothenia bernacchii* from Terra Nova Bay (Antarctica) during a five year period. Int. J. Environ. Anal. Chem. 61: 239-48.

Moreau C., Mah C., Agüera A., Améziane N., Barnes D., Crokaert G., Eléaume M., Griffiths H., Guillaumot C., Hemery L.G., Jażdżewska A., Jossart Q., Laptikhovsky V., Linse K., Neill K., Sands C., Saucède T., Schiaparelli S., Siciński J., Vasset N., Danis B. (2018) Antarctic and Sub-Antarctic Asteroidea database. Zookeys, 747: 141–156.

Negri A., Burns K., Boyle S., Brinkman D., Webster N., 2006. Contamination in sediments, bivalves and sponges of McMurdo Sound, Antarctica. Environmental Pollution 143: 456-467.

Nigro M., Orlando E., Regoli F., 1992. Ultrastructural localisation of metal binding sites in the kidney of the Antarctic scallop Adamussium colbecki. Marine Biology, 113: 637-643.

Nigro M., Regoli F., Rocchi R., Orlando E., 1997. Heavy metals in Antarctic Molluscs. In "Antarctic Communities" (B. Battaglia, J. Valencia and D.W.H Walton Eds.), Cambridge University Press, 409-412

Nonnis Marzano F., Fiori F., Jia G., Chiantore M., 2000. Anthropogenic radionuclides bioaccumulation in Antarctic marine fauna and its ecological relevance. Polar Biology 23: 753–58.

Olmastroni, S.; Ferretti, F.; Burrini, L.; Ademollo, N.; Fattorini, N. Breeding Ecology of Adélie Penguins in Mid Victoria Land, Ross Sea Antarctica. Diversity 2022, 14, 429. https://doi.org/10.3390/d14060429

Pane L., Feletti M., Francomacaro B., Mariottini G.L., 2004. "Summer coastal zooplankton biomass and copepod community structure near the Italian Terra Nova Base (Terra Nova Bay, Ross Sea, Antarctica)" Journal of Plankton Research, vol 26, issue 12, 1479-1488.

Papetti C., Babbucci M., Dettai A., Basso A., Lucassen M., Harms L., Bonillo C., Heindler F.M., Patarnello T., Negrisolo E. (2021). Not Frozen in the Ice: Large and dynamic rearrangements in the mitochondrial genomes of the antarctic Fish. Genome Biology and Eoluiton, 13–IS, DOI: 10.1093/gbe/evab017.

Peel S., Hill N., Foster S., Wotherspoon S., Ghiglione C., Schiaparelli S. (2019) Reliable species distributions are obtainable with sparse, patchy and biased data by leveraging over species and data types. Methods in Ecology and Evolution, 10: 1002-1014.

Piazza P., Cummings V., Guzzi A., Hawes I., Lohrer D., Marini S., Marriott P., Menna F., Nocerino E., Peirano A., Kim S., Schiaparelli S. (2019) Underwater photogrammetry in Antarctica: long-term observations in benthic ecosystems and legacy data rescue. Polar Biology, 42: 1061-1079.

Piazza P., Cummings V., Lohrer D., Marini S., Marriott P., Menna F., Nocerino E., Peirano A., Schiaparelli S., 2018. Divers-operated underwater photogrammetry: applications in the study of Antarctic benthos. The International Archives of the Photogrammetry, Remote Sensing and Spatial Information Sciences, Volume XLII-2, 885-892. https://doi.org/10.5194/isprs-archives-XLII-2-885-2018.

Piazza P., S.A. Gattone, A. Guzzi, S. Schiaparelli (2020) Towards a robust baseline for long-term monitoring of Antarctic coastal benthos, Hydrobiologia, 847: 1753-1771.

Piazza P., Błażewicz-Paszkowycz M., Ghiglione C., Alvaro M.C., Schnabel K., Schiaparelli S., 2014. Distributional records of Ross Sea (Antarctica) Tanaidacea from museum samples stored in the collections of the Italian National Antarctic Museum (MNA) and the New Zealand National Institute of Water and Atmospheric Research (NIWA). ZooKeys, 451: 49–60.

Povero P., Castellano M., Ruggieri N., Monticelli L.S., Saggiomo V., Chiantore M.C., Guidetti M., Cattaneo-Vietti R., 2006. "Water column features and their relationship with sediments and benthic communities along the Victoria Land coast, Ross Sea, Antarctica, summer 2004" Antarctic Science 18 (4), 603-613.

Povero P., Chiantore M., Misic C., Budillon G., Cattaneo-Vietti R., 2001. Pelagic-benthic coupling in Adélie Cove (Terra Nova Bay, Antarctica): a strongly land forcing controlled system? Polar Biology 24: 875-882.

Puce S., Cerrano C., Bavestrello G., 2002. *Eudendrium* (Cnidaria, Anthomedusae) from the Antarctic Ocean with a description of new species. Polar Biology 25: 366-73.

Pusceddu A., Cattaneo-Vietti R., Albertelli G., Fabiano M., 1999. Origin, biochemical composition and vertical flux of particulate organic matter under the pack ice in Terra Nova Bay (Ross Sea, Antarctica) during late summer 1995. Polar Biology 22: 124-32.

Regoli F., Nigro M., Benedetti M., Fattorini D., Gorbi S., 2005b. Antioxidant efficiency in early life stages of the Antarctic silverfish *Pleuragramma antarcticum*: Responsiveness to pro-oxidant conditions of platelet ice and chemical exposure. Aquatic Toxicology, vol. 75: 43–52.

Regoli F., Nigro M., Benedetti M., Gorbi S., Pretti C., Gervasi P.G., Fattorini D., 2005a. Interactions between metabolism of trace metals and xenobiotics agonist of the aryl hydrocarbon receptor in the Antarctic fish *Trematomus bernacchii*: environmental perspectives. Environmental Toxicology and Chemistry, vol. 24(6): 201–208

Regoli F., Nigro M., Bertoli E., Principato G.B., Orlando E., 1997b. Defences against oxidative stress in the Antarctic scallop *Adamussium colbecki* and effects of acute exposure to metals. Hydrobiologia, 355: 139-144.

Regoli F., Nigro M., Bompadre S., Wiston G., 2000a. Total oxidant scavenging capacity (TOSC) of microsomal and cytosolic fractions from Antarctic Arctic and Mediterranean Scallops: differentiation between three different potent oxidants. Aquatic Toxicology, 49: 13-25.



Regoli F., Nigro M., Chiantore M.C., Gorbi S., Wiston G., 2000b. Total oxidant scavenging capacity of Antarctic, Arctic and Mediterranean scallops. Italian Journal of Zoology, vol. 67: 5–94.

Regoli F., Nigro M., Chierici E., Cerrano C., Schiaparelli S., Totti C., Bavestrello G., 2004. Variations of antioxidant efficiency and presence of endosymbiontic diatoms in the Antarctic porifera *Haliclona dancoi*, Marine Environmental Research, vol. 58: 637-640.

Regoli F., Nigro M., Orlando E., 1998. Lysosomal and antioxidant defences to metals in the Antarctic scallop *Adamussium colbecki*. Aquatic Toxicology, 40: 375-392.

Regoli F., Principato G.B., Bertoli E., Nigro M., Orlando E., 1997a. Biochemical characterisation of the antioxidant system in the scallop *Adamussium colbecki*, a sentinel organism for monitoring the Antarctic environment. Polar Biology, 17: 251-25.

Regoli F., M. Nigro, M. Chiantore, G.W. Winston, 2002. Seasonal variations of susceptibility to oxidative stress in *Adamussium colbecki*, a key bioindicator species for the Antarctic marine environment. The Science of the Total Environment, 289: 205-211.

Regoli F., Nigro M., Chierici E., Cerrano C., Schiaparelli S., Totti C., Bavestrello G., 2004. Variations of antioxidant efficiency and presence of endosymbiotic diatoms in the Antarctic porifera *Haliclona dancoi*. Marine Environmental Research, 58: 637–640.

Rossi L., Sporta Caputi S., Calizza E., Careddu G., Oliverio M., Schiaparelli S., Costantini M.L. (2019) Antarctic food web architecture under varying dynamics of sea ice cover. Nature Scientific Reports, 9(1): 1–13.

Sarà A., Cerrano C., Sarà M., 2002. Viviparous development in the Antarctic sponge *Stylocordyla borealis* Loven, 1868. Polar Biology 25: 425-31.

Sarà M., Balduzzi A., Barbieri M., Bavestrello G., Burlando B., 1992. Biogeographic traits and checklist of Antarctic demosponges. Polar Biology 12: 559-85.

Schiaparelli S., Aliani, 2019. Oceanographic moorings as year-round laboratories for investigating growth performance and settlement dynamics in the Antarctic scallop *Adamussium colbecki* (E.A. Smith, 1902). PeerJ 7:e6373, DOI 10.7717/peerj.6373

Schiaparelli S., Linse K., 2006. A reassessment of the distribution of the common Antarctic scallop *Adamussium colbecki* (Smith, 1902). Deep-Sea Research II, 53: 912–920.

Schiaparelli S., Albertelli G., Cattaneo-Vietti R., 2003. The epibiotic assembly on the sponge *Haliclona dancoi* (Topsent, 1901) at Terra Nova Bay (Antarctica, Ross Sea). Polar Biology, 26: 342-347.

Schiaparelli S., Alvaro M.C., Kilgallen N., Scinto A., Lorz A.N., 2015. Host-shift speciation in Antarctic symbiotic invertebrates: further evidence from the new amphipod species *Lepidepecreella debroyeri* from the Ross Sea? Hydrobiologia 761: 143-159.

Schiaparelli S., Alvaro M.C; Barnich R., 2011. Polynoid polychaetes living in the gut of irregular sea urchins: a first case of inquilinism in the Southern Ocean and an overview of polychaete-echinoderm associations. Antarctic Science, 144-151 23 (2).

Schiaparelli S., Cattaneo-Vietti R., Chiantore M., 2000. Adaptive morphology of *Capulus subcompressus* Pelseneer, 1903 (Gastropoda: Capulidae) from Terra Nova Bay, Ross Sea (Antarctica). Polar Biology 23: 11-16.

Schiaparelli S., Ghirardo C., Bohn J., Chiantore M., Albertelli G., Cattaneo-Vietti R. 2007. Antarctic associations: the parasitic relationship between the gastropod *Bathycrinicola tumidula* (Thiele, 1912) (Ptenoglossa: Eulimidae) and the comatulid *Notocrinus virilis* Mortensen, 1917 (Crinoidea: Notocrinidae) in the Ross Sea. Polar Biology, 30: 1545–1555.

Schiaparelli S., Lörz A.N., Cattaneo-Vietti R., 2006. Diversity and distribution of mollusc assemblages on the Victoria Land coast and the Balleny Islands, Ross Sea, Antartica. Antarctic Science, 18 (4): 615–631.

Schiaparelli S., Cattaneo-Vietti R., Mierzejewski P., 2004. A "protective shell" around the larval cocoon of *Cephalodiscus densus* Andersson, 1907 Graptolithoidea (Hemichordata). Polar Biology, 27: 813-817.

Schiaparelli S., Alvaro M.C., Bohn J., Albertelli G., 2010. "Hitchhiker" polynoid polychaetes in cold deep waters and their potential influence on benthic soft bottom food webs. Antarctic Science, 399- 407 22 (4).

Schiaparelli S., Jirkov I.A., 2016A reassessment of the genus Amphicteis Grube, 1850 (Polychaeta: Amphaetidae) with the description of *Amphicteis teresae* sp. Nov. from Terra Nova Bay (Ross Sea, Antarctica). Italian Journal of Zoology 83: 531-542.

Schiaparelli, S., & Jirkov I. A. (2021) Contribution to the taxonomic knowledge of Ampharetidae (Annelida) from Antarctica with the description of *Amage giacomobovei* sp. nov.. European Journal of Taxonomy, 733: 125–145.

Schwaha T., Cometti V., Saadi A.J., Cecchetto M., Schiaparelli S. (2023) *Alcyonidium kuklinskii* sp. nov., a new species of Antarctic ctenostome bryozoan with a key to all Antarctic species of the genus. Organisms, Diversity & Evolution. https://doi.org/10.1007/s13127-023-00629-4.

Sciuto K., Moschin E., Alongi G., Cecchetto M., Schiaparelli S., Caragnano A., Rindi F., Moro I. (2021) *Tethysphytum* gen nov. and *Tethysphytum antarcticum* sp. nov. (Hapalidiales, Rhodophyta), a new non-geniculate coralline alga from Terra Nova Bay (Ross Sea, Antarctica): morpho-anatomical characterization and molecular phylogeny. European Journal of Phycology.

Signa G., Calizza E., Costantini, M.L., Tramati C., Sporta Caputi S., Mazzola A., Rossi L., Vizzini, S., 2019. Horizontal and vertical food web structure drives trace element trophic transfer in Terra Nova Bay, Antarctica. Environmental Pollution 246: 772-781.

Simeoni U., Baroni C., Meccheri M., Taviani M., Zanon G., 1989. Coastal studies in Northern Victoria Land (Antarctica): Holocene beaches of Inexpressible island, Tethys Bay and Edmonson Point. Boll. Ocean. Teor. Appl. 7(1-2): 5-16.

- (



Sterrer W., Sørensen M.V., Cecchetto M., Martínez A., Sabatino R., Eckert E.M., Fontaneto D., Schiaparelli S. (2022) First Record of the Phylum Gnathostomulida in the Southern Ocean. Diversity, 14, 382.

Stocchino C., Lusetti C., 1988. Le costanti armoniche di marea di Baia Terra Nova (Mare di Ross, Antartide). F.C. 1128 Istituto Idrografico della Marina, Genova.

Stocchino C., Lusetti C., 1990. Prime osservazioni sulle caratteristiche idrologiche e dinamiche di Baia Terra Nova (Mare di Ross, Antartide). F.C. 1132 Istituto Idrografico della Marina, Genova.

Swadling K.M., Penot F., Vallet C., Rouyer A., Gasparini S., Mousseau L., Smith M., Goffart A., Koubbi P., 2003. "Interannual variability of zooplancton in the Dumont d'Urville sea (39°E-146°E), east Antarctica, 2004-2008" Polar Science 5, 118-133, (2011)

Tolomeo A.M., Carraro A., Bakiu R., Toppo S., Garofalo F., Pellegrino D., Gerdol M., Ferro D., Place S.P., Santovito G. (2019) Molecular characterization of novel mitochondrial peroxiredoxins from the Antarctic emerald rockcod and their gene expression in response to environmental warming. Comp. Biochem. Physiol. C 255, 108580. https://doi.org/10.1016/j.cbpc.2019.108580

Tolomeo A.M., Carraro A., Bakiu R., Toppo S., Gerdol M., Irato P., Pellegrino D., Garofalo F., Bisaccia P., Corrà F., Ferro D., Place S.P., Santovito G. (2020) Too warm or not too warm... Is the antioxidant system of Antarctic fish ready to face climate changes? ISJ-Invertebr. Surviv. J., 17, 11. https://doi.org/10.25431/1824-307X/isj.v0i0.9-23

Trentin R., Moschin E., Duarte Lopes A., Schiaparelli S., Custódio L., Moro I. (2022) Molecular, Morphological and Chemical Diversity of two new species of Antarctic Diatoms, *Craspedostauros ineffabilis* sp. nov. and *Craspedostauros zucchellii* sp. nov.. Journal of Marine Science and Engineering, 10(11), 1656.

Trentin R., Moschin E., Grapputo A., Rindi F., Schiaparelli S., Moro I. (2023) Multi-gene phylogeny reveals a new genus and species of Hapalidiales (Rhodophyta) from Antarctica: *Thalassolithon adeliense* gen. et sp. nov. Phycologia. DOI: 10.1080/00318884.2022.2147745.

Tagliabue A. & Arrigo K.R., "Anomalously low zooplankton abundane in the Ross Sea: An alternative explanation" Limnol. Oceanogr. 48, 686-699.

Vacchi M., Cattaneo-Vietti R., Chiantore M., Dalù M., 2000a. Predator-prey relationship between nototheniid fish *Trematomus bernacchii* and Antarctic scallop *Adamussium colbecki* at Terra Nova Bay (Ross Sea). Antarctic Science 12(1): 64-68.

Vacchi M., Greco S., 1994a. Capture of the giant Nototheniid fish *Dissostichus mawsoni* in Terra Nova Bay (Antarctica): Notes on the fishing equipment and the specimens caught. Cybium 18(2): 199-203.

Vacchi M., Greco S., La Mesa M., 1991. Ichthyological survey by fixed gears in Terra Nova Bay (Antarctica). Fish list and first results. Memorie di Biologia Marina e di Oceanografia 19: 197-202.

Vacchi M., La Mesa M., 1995. The diet of Antarctic fish *Trematomus newnesi* Boulenger, 1902 (Notothenidae) from Terra Nova Bay, Ross Sea. Antarctic Science 7(1): 37-38.

Vacchi M., La Mesa M., 1997. Morphometry of Cryodraco specimens of Terra Nova Bay. Cybium 21(4): 363-68.

Vacchi M., La Mesa M., Castelli A., 1994b. Diet of two coastal nototheniid fish from Terra Nova Bay, Ross Sea. Antarctic Science 6(1): 61-65.

Vacchi M., La Mesa M., Greco S., 2000b. The coastal fish fauna of Terra Nova Bay, Ross Sea (Antarctica). In: Ross Sea Ecology, F. Faranda, L. Guglielmo and A. Ianora Eds., Springer Verlag, Berlin Heidelberg: 457-68.

Vacchi M., La Mesa M., Eastman J.T., 2004a."The role of notothenioid fish in the food web of the Ross Sea shelf waters: a review" Polar Biology 27(6), 321–338, (2004)

Vacchi M., La Mesa M., Dalù M., MacDonald J., 2004b. Early life stages in the life cycle of Antarctic silverfish, *Pleuragramma antarcticum* in Terra Nova Bay, Ross Sea. Antarctic Science

Vacchi M., Romanelli M., La Mesa M., 1992. Age structure of *Chionodraco hamatus* (Teleostei, Channichthyidae) samples caught in Terra Nova Bay, East Antarctica. Polar Biology 12: 735–38.

Van dijken G.L., Arrigo K.R., 2005. " Annual cycles of sea ice and phytoplankton in three Ross Sea polynyas" Poster at 3rd International Conference on the Oceanography of the Ross Sea Antarctica. Venezia, Italy, 10-14 Oct.



#### Map 1 Terra Nova Bay ASPA N° 161, Victoria Land, Ross Sea.

